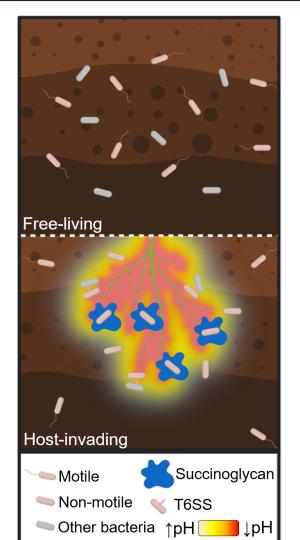
Succinoglycan Production in Agrobacterium tumefaciens

Amara Mason, Jacob Bouchier, and Pamela J.B. Brown Division of Biological Sciences, University of Missouri, Columbia MO



What is Succinoglycan?



Current model for plant invasion

Succinoglycan is a negatively charged polysaccharide that is required for Agrobacterium tumefaciens to invade a plant host. This invasion is the cause of Crown Gall disease, where the plant grows large tumor-like growths. The mechanistic role of succinoglycan in this process remains largely unknown.

Possible succinoglycan roles:

- Acid tolerance
- Osmoprotection
- Surfactant
- Signaling molecule

Calcofluor white staining reveals differential regulation of succinoglycan biosynthesis.

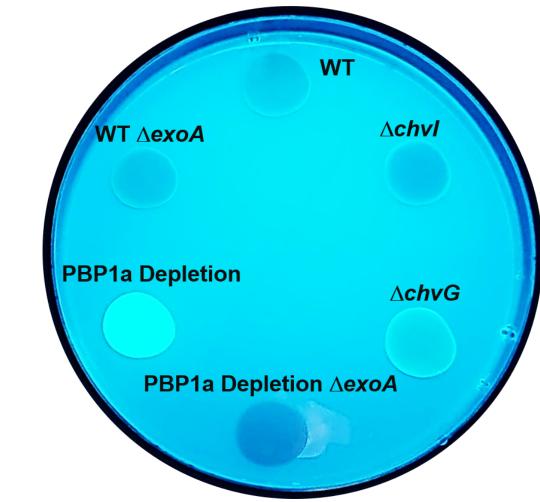


Image taken under UV light Greater fluorescence intensity = more succinoglycan produced

Preliminary results

- Cells that depleted of the cell wall synthase PBP1a have an overproduction of succinoglycan
- Δ*chvG* cells showed an unexpected production of succinoglycan, suggesting the existence of additional regulatory proteins.

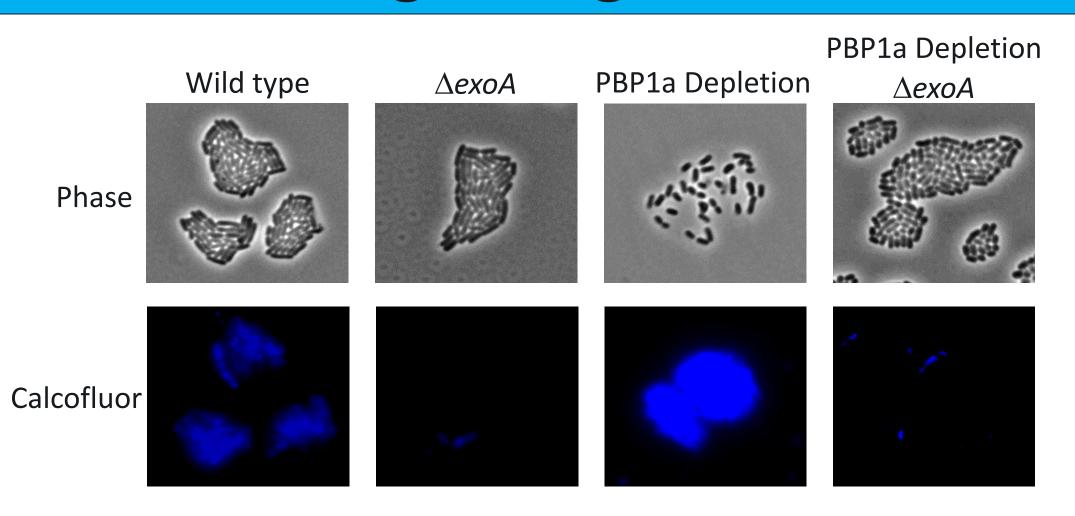
Objectives

Research Questions:

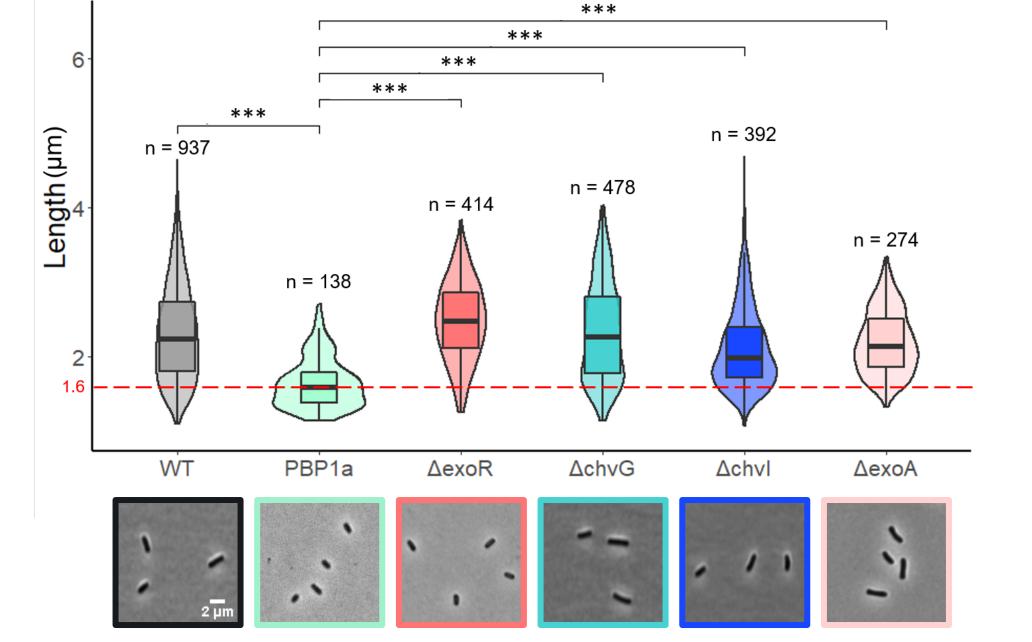
- Does succinoglycan production regulate growth?
- Does the impairment of growth signal for succinoglycan production?

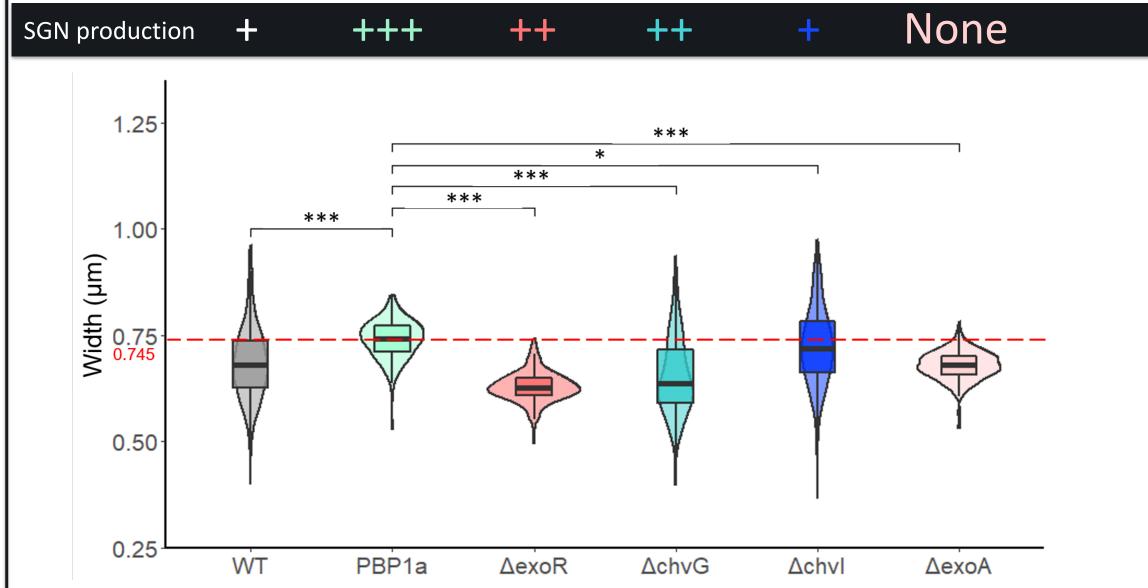
Hypothesis: Succinoglycan production is increased as a result of decreased growth.

Does succinoglycan production regulate growth?

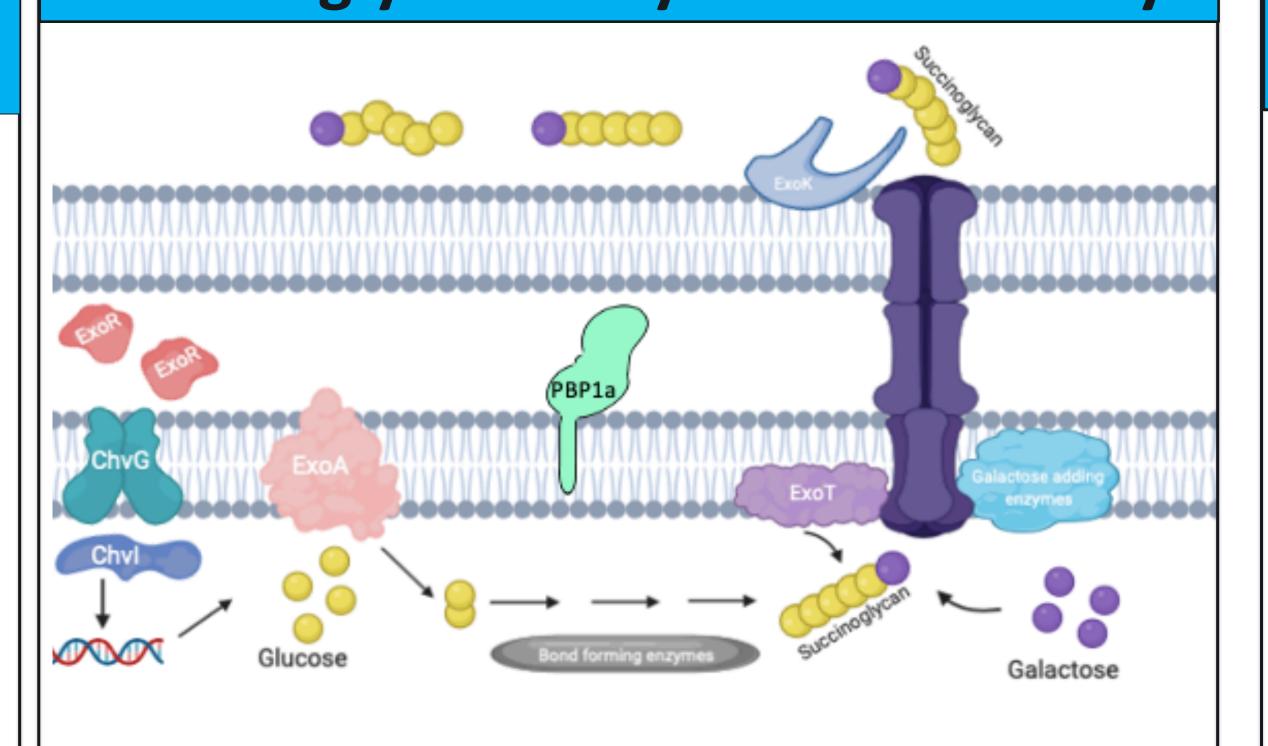


PBP1a-depleted cells are short and round, suggesting they are growth deficient. They also overproduce succinoglycan, suggesting a link between succinoglycan production and growth.



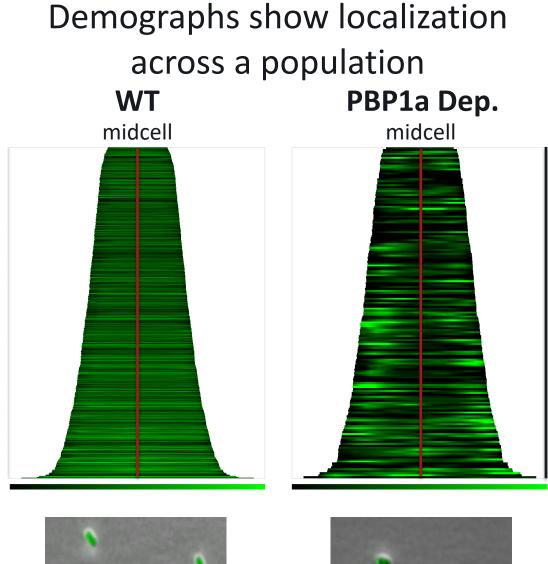


Succinoglycan Biosynthesis Pathway



How is succinoglycan secretion machinery coordinated in the cell?

ExoT-GFP fusion localizes the succinoglycan secretion complex.



Wildtype cells

grown for 16 hours

on an agarose pad

. PBP1a-depleted

cells grown for 16 hours liquid media **PBP1a Depletion**

PBP1a-depleted Wildtype cells cells grown for 16 grown for 16 hours on an agarose pad hours on an agarose

PBP1a Depletion After 36 **Hours of** growth After 72 hours of growth

Polar localization of ExoT-GFP appears dependent on either microcolony formation or PBP1a depletion

Development of a screen to detect succinoglycan regulatory mutants

1. A library of transposon insertion mutants will be generated using a method for random transposon mutagenesis.

2. Using the fluorescence intensity readout of calcofluor bound to succinoglycan, the library will be screened for mutants that no longer overproduce succinoglycan when depleted of PBP1a.

3. PBP1a expression will be controlled using a chemical inducer known as IPTG.

4. Mutants will be classified and then sequenced for further characterization.

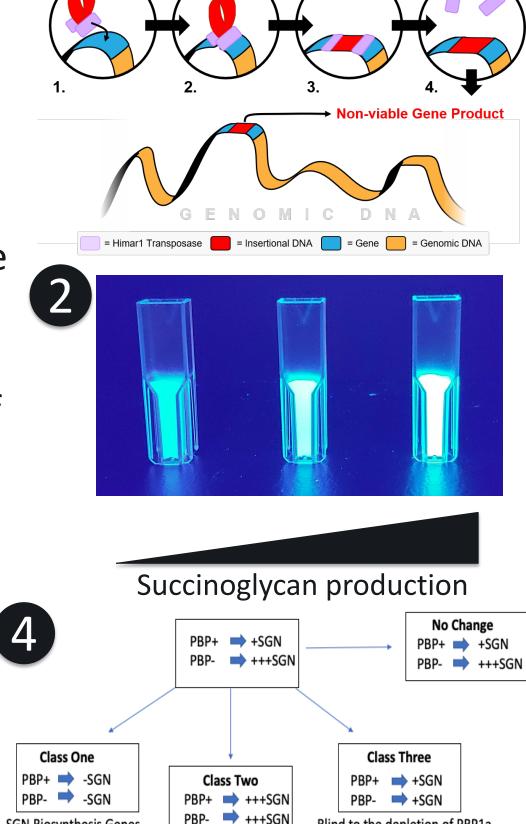
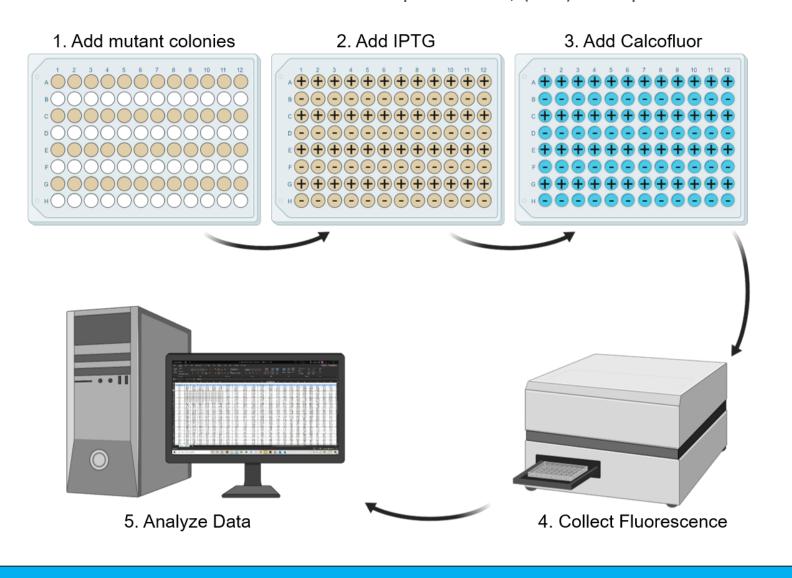


Diagram showing classes of data we expect to find when running our screen. (-) No production; (+) Some production; (+++) Over production. SGN = Succinoglycan.



Acknowledgements

Thank you to Dr. Brown and the Brown lab from providing me with this valuable experience of conducting research. I would like to give a sincere thank you to my graduate student mentor, Jacob Bouchier, for endlessly supporting and assisting me throughout this journey. Funding: University of Missouri Maximizing Access to Research Careers Program (T34)