

# SEXUAL DIMORPHISM IN NEURON COUNT AND DENSITY IN ANOLIS CRISTATELLUS

Jessica Garcia<sup>1</sup>, Levi Storks<sup>2</sup> and Manuel Leal<sup>2</sup>

1. University of Missouri; College of Arts and Sciences 2. University of Missouri; Division of Biological Sciences

# Background

- Adaptive radiation of the Anolis in the Greater Antilles has resulted in many different *Anolis* species that are morphologically and behaviorally differentiated based on the environment that they occupy (1). It has been suggested that sexual dimorphism plays an important role in ecomorphological diversity due to sexual variation in morphology, habitat preference, diet and behavior (2).
- Previous studies of Anolis brain have primarily focused on the relationship between relative brain size in relation to structural habitat (3, 4). However, the link between sexual size dimorphism and neuronal density is poorly understood, even though there is a distinct difference in brain size.

# Objective

**Evaluate if sexual size dimorphism impacts neuronal** density in Anolis cristatellus.

#### Methods

- Anoles were intracardially perfused using PBS, followed by 4% paraformaldehyde.
- ❖ The brains were dissected into three regions (cerebellum, telencephalon and rest of brain), and weighed.
  - The rest of brain consists of the diencephalon, mesencephalon, pons and myelencephalon.

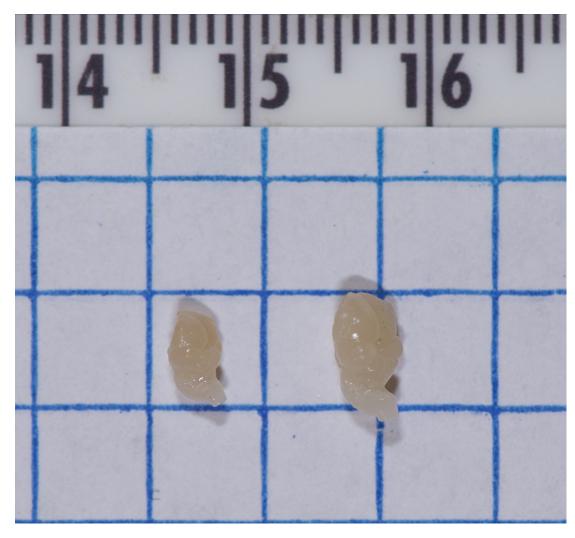


Figure 1: comparison of female (left) and male *Anolis* cristatellus brains.

- Each brain region was put into a 1mL tissue grinder with dissociation solution (1% Triton-X + 40mM sodium citrate) and homogenized for at least 15 minutes.
- The homogenate was collected using a Pasteur pipette into a 15mL centrifuge tube after repeated rinsing.
- The homogenized tissue was centrifuged to isolate the nuclei pellet and remove supernatant. The tube volume was adjusted with PBS to 10<sup>6</sup> nuclei/mL.
- ❖ DAPI was added to the tubes at a 20x dilution (2).
- The tube was inverted, and samples were aliquoted onto the hemocytometer and counted under the Leica DM5500 fluorescence microscope.
  - Four or more samples were counted or until coefficient of variance was 0.15 or lower to ensure consistency.
  - Average of counts was calculated from these four values.
  - Erythrocyte nuclei were excluded from count by their shape and their autofluorescence.



Figure 2: Image showcasing size difference between adult male (upper) and female (lower) Anolis cristatellus

### Results

**Table 1**: Cell counts and region mass across the brain of female and male *Anolis cristatellus* 

Sex:	Mean Body Mass (g)	Region:	Mean Region Mass (mg):	Mean N x10 <sup>6</sup> :	Mean DN (cells/mg ) x10 <sup>6</sup> :
A. cristatellus male	4.6 ± 0.346	BR (n=9)	0.0457 ± 0.00154	4.87 ± 0.299	$0.107 \pm 0.007$
		TEL (n=10)	0.0168 ± 0.0071	$1.83 \pm 0.080$	0.11 ± 0.0069
		CER (n=9)	0.0017 ± 0.00007	$0.753 \pm 0.073$	$0.450 \pm 0.038$
		ROB (n=10)	0.0269 ± 0.00075	2.26 ± 0.178	0.084 ± 0.0068
A. cristatellus female	1.67 ± 0.133	BR (n=6)	0.0314 ± 0.00181	4.03 ± 0.159	0.131 ± 0.012
		TEL (n=6)	0.0117 ± 0.00094	$1.37 \pm 0.097$	0.122 ± 0.016
		CER (n=6)	0.0015 ± 0.00009	$0.659 \pm 0.039$	$0.454 \pm 0.047$
		ROB (n=6)	0.0182 ± 0.00080	$2.00 \pm 0.100$	0.111 ± 0.0084

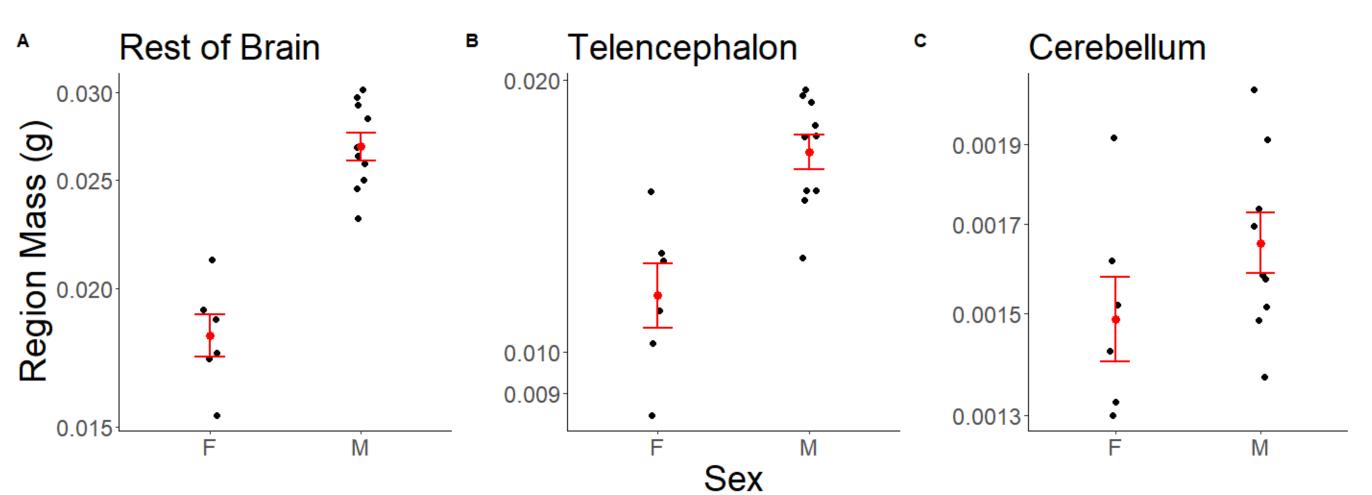


Figure 3: Males and females exhibited differences in region mass in the rest of brain (p>1.68 x 10<sup>-6</sup>), telencephalon (p>0.00075), but not the cerebellum (p<0.15). The red dot represents the mean value. The bars represent standard error. The data points are jittered.

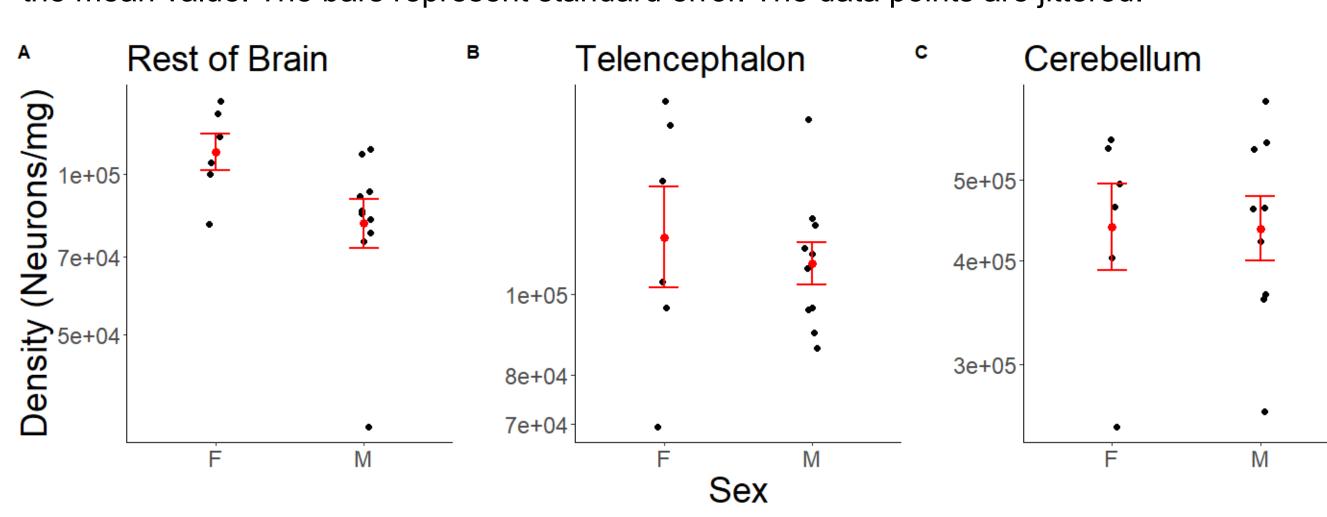
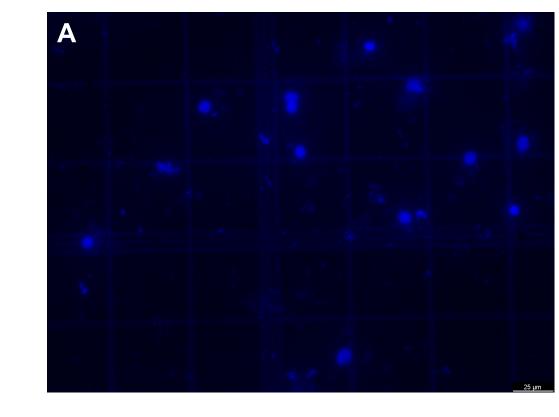


Figure 4: Males and females trended toward significance in neuronal density in the rest of brain (p<0.06) but not in the telencephalon (p<0.59) and cerebellum (p<0.98). The red dot represents the mean value. The bars represent standard error. The data points are jittered.

#### **Methods Continued**

- ❖ The total number of cells in the solution was then calculated
- $\frac{Cells\ counted}{0.0001\ mL} \times \text{total volume of solution (mL)} = \text{total cells in solution}$
- ❖ The samples were immunoreacted to label neuronal nuclei (NeuN; ABN78C3) and then counted.
  - We counted the proportion of labeled neurons out of 500 nuclei.
- Once the percent of neurons was obtained, that number was multiplied by the total number of cells
- Values were compared between males and females using ANOVA



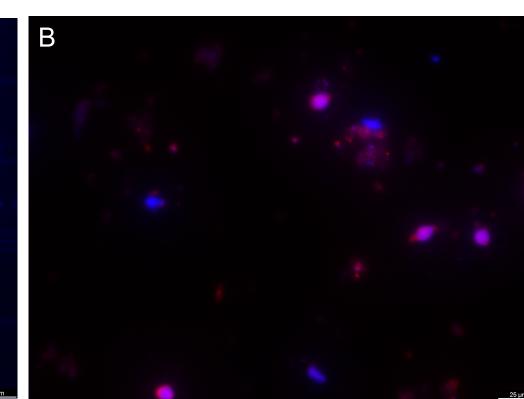


Figure 5: A) Image at 400x magnification showing brain nuclei stained with DAPI within the gridlines of the hemocytometer. B) Image at 400x magnification showing brain nuclei double labeled with DAPI (blue) neuronal nuclei antibody (red)

## Conclusion

- Our results show that there is a difference in total brain mass, telencephalon and rest of brain region mass, but not in the cerebellum between male and female *Anolis* cristatellus.
  - The differences in brain mass between sexes is likely due to the differences in body mass.
  - There is not a significant difference in cerebellar mass between males and females
    - Could be due to less variation in cerebellum size as they get bigger in body size
- There is no significant difference between neuronal density in male and female *Anolis* in the telencephalon or cerebellum. However, there is a trend toward significant difference in neuronal density in the rest of brain.
- Males and females might require the same density of neurons in the telencephalon and cerebellum
- Females might require larger neuronal density in rest of brain than males

# **Next Steps**

- Collecting more brains to increase our sample size and see if the same trends hold true with more data.
- Looking at the neuronal density of different Anolis ecomorphs, and see if different degrees of sexual size dimorphism influences neuronal density

- Indian Anolis Lizards." Animal Behaviour, vol. 75, no. 2, Feb. 2008, pp. 555-63.
- 3. Powell, Brian J., and Manuel Leal. "Brain Organization and Habitat Complexity in Anolis Lizards.\_" Brain, Behavior and Evolution, vol. 84, no. 1, 2014, pp. 8–18.
- 4. De Meester, Gilles, et al. "Brain Size, Ecology and Sociality: A Reptilian Perspective." Biological Journal of the Linnean Society, vol. 126, no. 3, Feb. 2019, pp. 381–91 5. Herculano-Houzel, S., von Bartheld, C.S., Miller, D.J., Kass, J.H. 2015. "How to Count Cells: The Advantages and Disadvantages of the Isotropic Fractionator Compared
- with Stereology." Cell and Tissue Research, vol. 360, no. 1, Apr. 2015, pp. 29–42.

#### **Acknowledgements:**

I would like to take time to thank members of the Chipojo lab for going to Puerto Rico to collect the lizards used in this study. Funding for my undergraduate research came from Initiative for Maximizing Student Diversity Fellows Program via grant number R25GM056901 from the National Institute of General Medical Science (NIGMS), a component of the National Institutes of Health (NIH). I would also like to thank the University of Missouri Cytology Core for letting me use the Leica DM5500 fluorescence scope in order to obtain the neuronal and nonneuronal counts and data.

<sup>1.</sup> Butler, M.A., Sawyer, S.A., Losos, J.B. 2007. "Sexual Dimorphism and Adaptive Radiation in Anolis Lizards." Nature, vol. 447, no. 7141, May 2007, pp. 202–05. 2. Johnson, M.A., Leal, M., Schettino, L.S., Lara, A.C., Revell, L.J., Losos, J.B. 2007. "A Phylogenetic Perspective on Foraging Mode Evolution and Habitat Use in West